

Basic Topspin commands for NMR at the BNSP

Mar-1-2018

Action	Command [‡]	Comment
Samples		
display lock window	lockdisp	adjust lock gain so lock is 80%
eject sample	ej	
edit temperature	edte	enter target temperature
inject sample	ij	check temperature before this!
lock	lock	[90% H ₂ O/10% D ₂ O] or [D ₂ O] or [Methanol-d4]
optimize lock feedback loop	loopadj	loopadj N where N is from 1-10 gives slow to fast reaction time
automatic tune/match module	atma	option exact slower but more accurate
manual tune/match module	atmm	adjust both tune & match to shift curve from left to right last to avoid drifting
read shims	rsh *.eth	read a recent shim
automatic topspin shimming	topshim gui	topshim (runs without opening gui)
manually adjust lockphase & shims	bsmsdisp	software version of the lock panel
Experiments		
edit current experiment	edc	NAME EXPNO PROCNO DIR /opt/topspin/ user /nmr change user if copying parameters from another user Do not select checkbox “Keep parameters”!
read parameters	rpar *.all	can filter e.g: rpar NA_*.all also switch from bruker to user dir!
write processed & acquired data	wrpa	can overwrite nmr data!
edit pulsesequence parameters	ased	calculated parameters are shown in grey
edit acquisition parameters	eda	includes parameters for indirect dimensions of 2D,3D
edit processing parameters	edp	
go set up adjust parameters during measurement	gs	adjust o1 , rg , flipback pulses interactively
display cnst array	cnst	all constants
display power level array	p1	all hard power levels
display pulse length array	p	all pulse lengths (in us)
display shaped pulse power array	spdb	all shaped pulses (name, power, etc)
zero go (measure data)	zg	delete current experiment buffer and start experiment
Calibrations		
temperature calibration	calctemp	first process methanol spectrum
Calibrations		
90° hard pulse calibration	Hcn90	enter logbook power levels to calibrate afterwards write calibrations in logbook!
set up DOTALL experiment	xxx90 geteth	set 90° hard pulses (p1,p3,p5 & pldb 1-3) set params e.g. shape pulse & dec pwr
set up Bruker experiment	getprosol 1H p1 pldb1 13C p3 pldb2 15N p5 pldb3	

Standard high power pulses			
Channel	Power level	90° pulse	180° pulse
1: H1	p1db1	p1	p2
2: C13	p1db2	p3	p4
3: N15	p1db3	p5	p6
3: N15 (Bruker pulseq.)	p1db3	p21	p22
See logbook for standard values			

Action	Command [‡]	Comment
Processing data		
exponential multiply	em	lb is linebroadening in Hz
sine window multiply	sin	phaseshifted by π/ssb but $ssb=0$ is unshifted sine)
quadratic sine window multiply	qsin	see above
fourier transform 1D	ft fp ef efp qfp	fourier transform only ft+pk em+ft em+ft+pk qsin+ft+pk
phase correction (interactive)	.ph	.sret (store phases, end phasing mode)
apply phase correction	pk	apply phc0 and phc1 to spectrum
automated phase correction	apk	only for simple spectra like methanol
Baseline correction	abs	absf1 & absf2 are range in ppm
2D spectra		
display params of indirect dim	eda	displays sw , td , offsets o1 in all dims td1 (# of fids) must be an even number!
fourier transform 2D	xfb xfb n xf2 xf1	FT both dimensions xfb and discard imaginary data FT direct dim (F2) FT indirect dim (F1)
Baseline correction	abs2 abs1	baseline correct F2 dimension baseline correct F1 dimension
3D spectra		
Fourier transform	tf3 n;tf2 n;tf1 n ftnd 0 ftnd 0 dlp	FT dim F3,F2,F1 n =no imaginary data FT all dims in AQORDER FT all dim with delayed linear pred. (dlp)
Baseline correction	tabs3;tabs2;tabs1	Baseline correct dim F3,F2,F1
Extract data		
Extract fid from 2D ser file	rser N M	read fid N and store in expno=M
read serial row	rsr N M	read row N and store in procno=M
read serial column	rsc N M	read column from processed 2D
If M is not included: fid/row/column is transferred to ~TEMP/1, overwriting what is there		

[‡]Many additional standard topspin commands are available as documented in the acquisition and processing manuals available from the Topspin's help button, or the BNSP website. In addition, there are a number of BNSP specific commands documented on the BNSP website. A few are given in the above table: **HCN90**, **xxx90**, **geteth**

Common procedures

Calibrate temperature

1. Create new experiment (**edc**)
2. Load parameter-set for temperature calibration (**rpar methanol4.eth**)
3. eject sample (**ej**)
4. insert methanol4.eth sample with yellow label (**ij**)
5. lock on methanol (**lock**) select [methanol-d4]
6. set temperature (**edte**)
7. read a recent shimfile (**rsh *.setup**)
Steps 8-12 can be performed on one line: **atma;topshim;zg;ef;apk;calctemp**
8. tune/match (**atma**)
9. shim (**topshim**)
10. measure data (**zg**)
11. process 1D (**ef;apk**)
12. calculate temperature (**calctemp**)
13. Adjust target temperature in edte and wait a few minutes
14. Repeat steps 7 to 10 until calibrated temperature = desired temperature
 - Now you can use this set temperature for all your samples
 - Temperature may still deviate from your desired value if experiment heats (e.g. decoupling, TOCSY)

Insert sample and set up 1D ¹H measurement

1. Create a new experiment (**edc**)
2. load experiment parameters (**rpar *.eth**) select zgpr.eth or zg-wg3919.eth
3. lock display (**lockdisp**)
4. eject sample (**ej**)
5. set temperature (**edte**)
6. insert sample (**ij**)
7. lock on H₂O/D₂O (**lock**)
8. optimize lock parameters for sample (**loopadj**)
9. tune and match probe (**atmm**)
10. read shims (**rsh *.eth**)
11. adjust lock or shims manually (**bsmsdisp**)
12. start automatic shimming (**topshim gui**)
13. set power levels: **p1db1** (¹H), **p1db2** (¹³C), **p1db3** (¹⁵N)
14. pulse length determination (**HCN90**)
 - If it fails due to low s/n, increase **ns**
 - After running **HCN90**, calibrate high pwr pulses for DOTALL or ETH datasets: **xxx90**
15. check and optimize experimental setup (**ased**)
16. interactively adjust parameters (**gs**)
 - Optimize **o1** for solvent suppression
 - Optimize **rg** to be large, but avoid receiver overflow
17. record a spectrum (**zg**)
18. stop experiment (if lock drops rapidly stop measurement: **stop**)
19. end experiment early after completing current phase cycle: **halt**)
20. process the spectrum (1D: **efp**, 2D: **xfb**)
21. phase correct if necessary (**.ph** to start, **.sret** to store results)

Calibrate selective pulses for solvent flip-back

1. Calibrate hard 90deg ¹H pulse using HCN90 (see procedure above)
On 900 do this manually use zg experiment to measure 360° pulse & divide by 4.
2. Create a new experiment (**edc**)
3. Read flipback calibration parameter-set (**rpar flips.all**)
4. Enter hard 90deg ¹H pulse calibration (**xxx90**)
5. Display important parameters and adjust them (**ased**)
 - p11 flipback pulselength – set to value appropriate for spectrometer
 - o1 – set to optimized value determined in zgpr .eth experiment using **gs**
 - ZGOPTNS :

Type of flip pulse	ZGOPTN	Shape name *	Shape power	Phase correction
flip down	-DDWN	spnam1	spdb1	phcor1
flip up	-DUP	spnam2	spdb2	phcor2
flip watergate	-DWG	spnam3	spdb3	phcor3

* Use gauss128_5, Sinc1000, or rect1000 and geteth for *approximate* calibration
Other shapes are possible, but then you must use **stdisp** for *approximate* calibration.

6. Calibrate shaped pulse powers to *approximate* values (**geteth**)
7. Interactively adjust shaped pulse pwr & phase correction to minimize solvent (spdb1, phcor11) (**gs**)
8. Write down the shaped pulse power and phase correction
9. Change to next type of flipback pulse (ZGOPTN: -DDWN, -DUP, DWG) and repeat steps 7-8

Setup and measure 2D (for bruker expt replace steps 3-4 with getprosol – see bottom of p.1)

1. Create a new experiment (**edc**)
2. Read 2D parameter-set (**rpar *.all**) and select 2D: E.g. HSQC15N .all
3. Enter high power pulse calibrations (**xxx90**, uses calibration from 1D above)
4. Setup 2D experiment automatically (**geteth**, calibrates decoupling, shapes, td1, td2)
5. Check main acquisition parameters (**ased**)
6. Check indirect dim acquisition parameters (**eda**)
 - Important are:
 - decoupling should not be too high ~4W, for 123ms
 - No negative calculated delays
 - Indirect dim carrier (**o3p** for experiments with 15N in indirect, **o2p** for 13C)
 - Indirect spectral width in ppm (**sw**) F1
7. Run 2D (**zg**)
8. Extract fid from 2D series (**rser N**) where **N** is fid number
 - FID is copied to 1D parameterset in ~TEMP/1
 - You can process this like the 1D described above, and store phases to 2D
 - Close window when done using x in upper right corner
9. Process direct (¹H) dimension of 2D (**xf2**)
10. Process indirect (¹⁵N) dimension (**xf1**)
11. Process both dimensions (**xfb**)
12. After initial 2D run with small number of scans (ns), adjust
 - **sw** (F1) and **o3p** (¹⁵N) or **o2p** (¹³C) to optimize spectral width to sample

Setup and measure 3D:

1. Steps 1-8, same as for 2D
2. process a 2D plane of 3D (with no evolution in one dim (**xfb**))
3. select orientation 13 or 23, and processing number 13 or 23
4. phase as usual for a 2D and store to 3D, close 2D by clicking on x at top right
5. revise **STSI** and **STSR** so strip FT selects region of interest in direct dim3
6. Enter **STSI** and **STSR** from step 2c above in 3D processing parameters
7. Process direct (¹H) dim (**tf3 n**)
8. Process indirect dim (**tf1 n** or **tf2 n**)
9. Process other indirect dim (**tf2 n** or **tf1 n**)
10. Steps 2-4 can be replaced by **ftnd N** or **ftnd 0 dlp**
11. The 3D can be viewed with 3D viewer after steps 4-7 to inspect results