Supplementary Material

Structure, Folding and Stability of FimA, the Main Structural Subunit of Type 1 Pili from Uropathogenic *Escherichia coli* Strains

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| FimAw | t | | | | | |
|-------------|----------|-------------|-----------|-------------|------------|----------------|
| 1 | 10 | 20 | 30 | 40 | 50 | 60 |
| AATTVN | GGTVHFK | GEVVNAACAVD | AGSVDQTVQ | LGQVRTASLAÇ | EGATSSAVG | FNIQLND |
| | 70 | 80 | 90 | 100 | 110 | 120 |
| CDTNVA | SKAAVAF | LGTAIDAGHTN | VLALQSSAA | GSATNVGVQII | DRTGAALTLI | GATFSS |
| | 130 | 140 | 150 | | | |
| ETTLNN | GTNTIPF | QARYFATGAAT | PGAANADAT | FKVQYQ | | |
| | | | | | | |
| | | | | | | |
| FimAa | | | | | | |
| 1 | 10 | 20 | 30 | 40 | 50 | 60 |
| AATTVN | GGTVHFK | GEVVNAACAVD | AGSVDQTVQ | LGQVRTASLAÇ | EGATSSAVG | FNIQLND |
| | 70 | 80 | 90 | 100 | 110 | 120 |
| CDTNVA | SKAAVAFI | LGTAIDAGHTN | VLALQSSAA | GSATNVGVQII | DRTGAALTLI | GATFSS |
| | 130 | 140 | 150 | 160 | 170 | 180 |
| ETTLNN | GTNTIPF | QARYFATGAAT | PGAANADAT | FKVQYQGGGGG | GAATTVNGG | <u>rvhfkge</u> |
| | | | | | | |
| <u>VVNA</u> | | | | | | |
| | | | | | | |
| E:ma+ | | | | | | |
| f TIIIWC | 10 | | ~~ | | 50 | C 0 |
| T | TO | 20 | 30 | 40 | 50 | 00 |
| | МНННННК | JEVVNAACAVD | AGSVDQTVQ | LGQVRTASLAQ | EGATSSAVG | SUTOTUD |
| ~~~~~ | 70 | 80 | 90 | 100 | 110 | 120 |
| CDTNVA | SKAAVAF | | VLALQSSAA | GSATNVGVQII | DRTGAALTLI | JGATESS |
| | 130 | 140 | 150 | | | |
| ETTLNN | GINTIPF | JARYFATGAAT | PGAANADAT | ŀKVQYQ | | |

Supplementary figure 1:

Amino acid sequences of FimAwt, FimAa and FimAt. The N-terminal (His)₆ tag in FimAt and the (Gly)₆-linker between the natural FimA C-terminus and the engineered, C-terminal donor strand in FimAa are indicated in italics, the C-terminal donor strand in FimAa is underlined. The amino acid numbering is according to the sequence of mature FimAwt.



Supplementary figure 2:

Far-UV CD spectra of FimAwt (a), FimAa (b) and FimAt (c) measured right after dilution (1:20) of unfolded protein (in 6 M GdmCl) with 10 mM sodium phosphate, pH 7.0, 200 mM NaCl buffer with (red) or without (blue) 6 M GdmCl. The measurement of one spectrum took less than three minutes. The black lines represent the corresponding spectra after several days of incubation in refolding buffer.



Supplementary figure 3:

Frequency of reported folding rates of model proteins. 104 rate constants are covered in this plot (55 folding rates of two-state folders, 39 rates of formation of the native state and 10 rates of intermediate formation for non-two-state folders).³⁸⁻⁴⁴ The folding rate of FimA is indicated by the red bar. The solid blue line represents a fit according to a Gaussian function (fit does not include FimA).

Table S1:

Contact order (CO), absolute contact order (ACO), and the number of sequence distant native pairs (Q_d) describing the topological complexity of the FimAa structure[#].

| | Value |
|-----------------------------------------------|--------|
| CO (averaged over all structures) | 13.5 % |
| CO (contacts present in all structures) | 18.2 % |
| ACO (averaged over all structures) | 22.6 |
| ACO (contacts present in all structures) | 30.4 |
| Q _d (averaged over all structures) | 201 |
| Q_d (contacts present in all structures) | 145 |

[#] Since NMR structures are represented by a number of slightly different conformers representing the structure it is not straightforward to calculate the contact parameters. We used two approaches: calculation of the parameter for all conformers individually and take its average, as well as considering only those contacts that are present in all structures.