

Characterization of Catalysts and Surfaces

Elemental Analysis (ICP, AAS etc.)

Fall Semester 2016

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HCI G105

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Outline

- Instrumental Methods for Determination of the Elements
 - Atomic Absorption Spectrometry - **AAS**
 - Inductively Coupled Plasma Optical Emission Spectrometry - **ICPOES**
 - Inductively Coupled Plasma Mass Spectrometry - **ICPMS**
- Sample Preparation Techniques
- Considerations for Quantitative Analysis

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Common Aspects

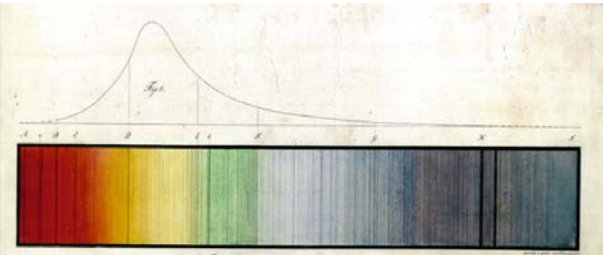
- Solution-based Techniques
 - Usually aqueous
 - Usually acidified (HNO_3 , HCl ; 1-10 Vol.%)
 - Digestion of solids required
- Calibration required
 - External standards / Standard Addition
 - Matrix-matching may be required
 - Internal standards (when applicable)

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Atomic Absorption Spectrometry



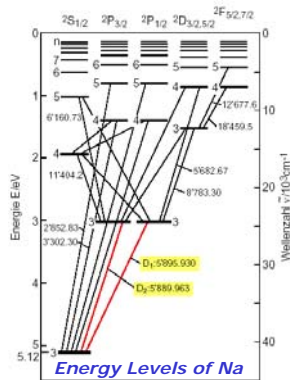
Gaseous atoms can absorb electromagnetic radiation corresponding to the ΔE of their electronic states

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Atomic Absorption

- Photons of sufficient energy excite electrons to higher levels
- Excitation is most likely for "resonant" photon, i.e.: $h\nu = \Delta E$



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Atomic Absorption Spectrometry

The magnitude of this absorption is given by **Lambert-Beer's law**:

$$I = I_0 e^{-\epsilon_\lambda Lc}$$

$$E = \lg \frac{I_0}{I} = \epsilon_\lambda Lc$$

- I: Intensity registered in presence of an absorber (analyte atom)
- I_0 : Intensity registered in absence of an absorber (analyte atom)
- ϵ_λ : Absorption coefficient
- λ : Wavelength registered
- L: Observation path length
- c: Concentration of the analyte
- E: Extinction (evaluated signal)

AAS (and ICPOES) operate in the UV-VIS range, $\approx 200 - 800$ nm

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Atomic Absorption Spectrometry

So what do we need for an AAS Experiment?

- 1) Gaseous Atoms → Flame or Furnace
- 2) Photon source → line- or continuous
(ideally resonant $h\nu$) (HCL, EDL or arc)
- 3) Optical Spectrometer
(Monochromator, Detector)

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Atomic Absorption Spectrometry



Flame Atomizer

Oxidants: Air, N_2O , O_2
Fuel: Acetylene, H_2 , CH_4 , C_3H_8

Oxidant	Fuel	T, °C
Air	CH_4	1875
Air	C_2H_2	1920
Air	H_2	2045
Air	Acetylene	2300
N_2O	Acetylene	2750
O_2	H_2	2860
O_2	Acetylene	3100



Furnace Atomizer

Electrically heated up to 2700 °C

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Atomic Absorption Spectrometry

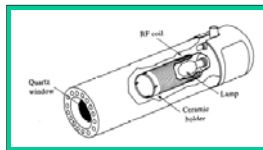
Line Sources

Element-specific emission spectra

Hollow Cathode Lamp
HCL

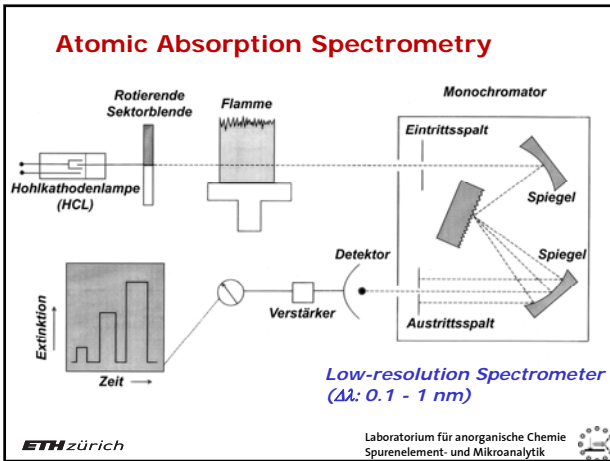


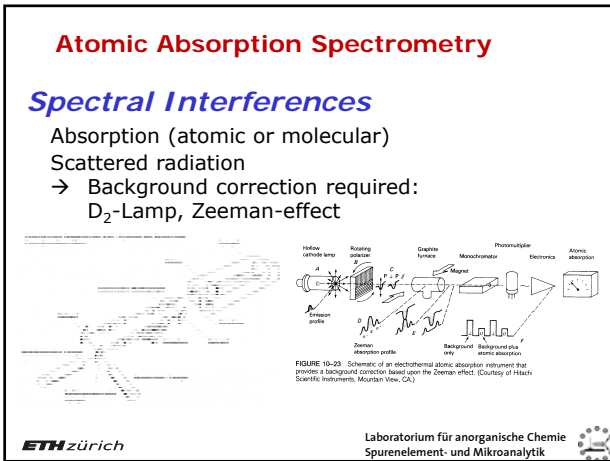
Electrodeless Discharge
Lamp
EDL

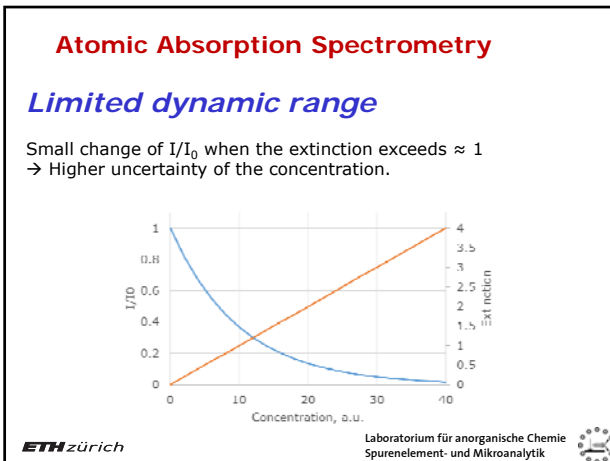


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Atomic Absorption Spectrometry

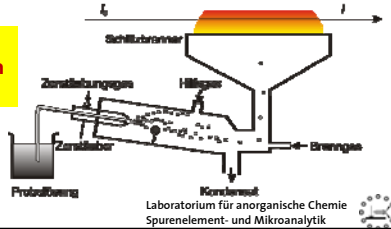
Flame AAS Principle

Sample is nebulized and aerosol transferred to burner.

Optimization:

- Fuel/Oxidant gas flow rates
- Observation height

Temperature
Particle formation
Residence time



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Atomic Absorption Spectrometry

Furnace AAS

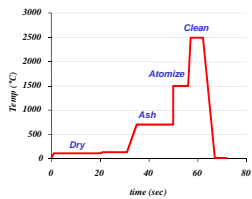


Sample is pipetted into furnace (addition of modifier) and T-program started.

Optimization:

- T-program: Drying, Ashing, Atomization, Cleaning
- Modifier type and amount

Separation of Analyte
from sample matrix and
Solvent.
Efficient vaporization.



Common Modifiers:
 $Mg(NO_3)_2$, $Pd(NO_3)_2$, $NH_4H_2PO_4$

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Atomic Absorption Spectrometry

Characteristics

	Flame AAS	Furnace AAS
Sample Volume	mL	μ L
Analysis Time	seconds	minutes
Calibration ¹	Ext./Std.Add.	Std.Add.
Limits of Detection ²	0.1 – 1000 μ g/L	0.001 -10 μ g/L
Repeatability ³	1-5%	1-5%

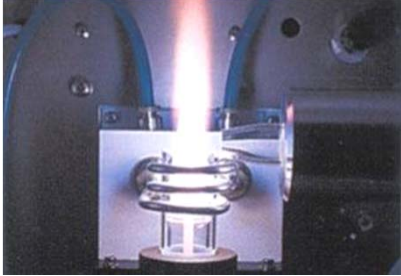
1: Ext: External Calibration, Std.Add.: Standard Addition
2: Depends on Element, Matrix, Analysis Time
3: For Concentrations > 100 \times LOD

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ICP-Optical Emission Spectrometry

ICP: Inductively Coupled Plasma
An RF-powered electrodeless gas (Ar) discharge,
reaching Temperatures of 10000K



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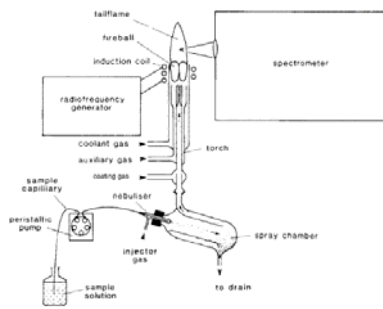
ICP-Optical Emission Spectrometry

Setup

Nebulizer Gas: ≈ 1 L/min Ar
Auxiliary Gas: ≈ 1 L/min Ar
Coolant Gas: ≈ 14 L/min Ar

RF-Power: 1000 - 1500 W
RF Frequency: 27, 40 MHz

Sample uptake: ≈ 1 mL/min
Nebulizer Efficiency: 3 - 5 %



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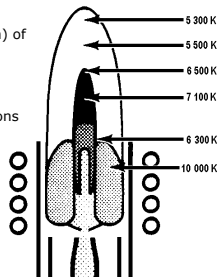
ICP-Optical Emission Spectrometry

Characteristics

Sample aerosol is injected in a narrow region (≈ 2 mm) of very high Temperature:

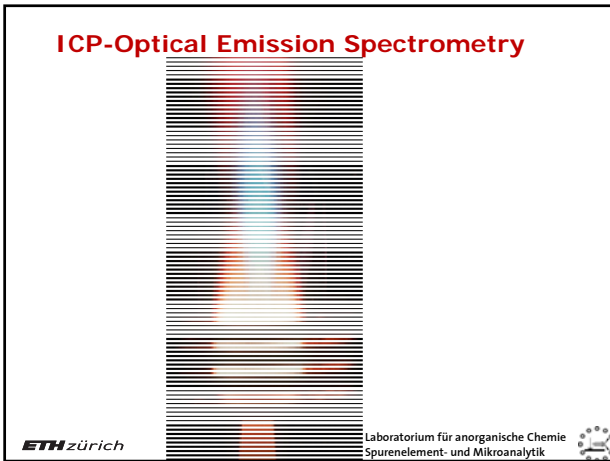
- Efficient atomization
- Ionization
- Generation of electronically excited Atoms and Ions

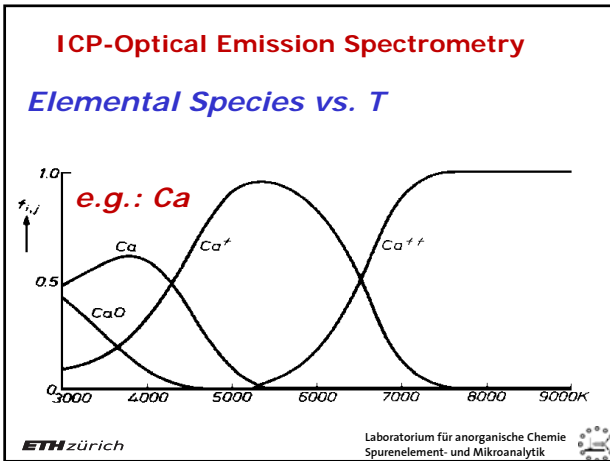
Relaxation of excited states leads to detectable emission signals

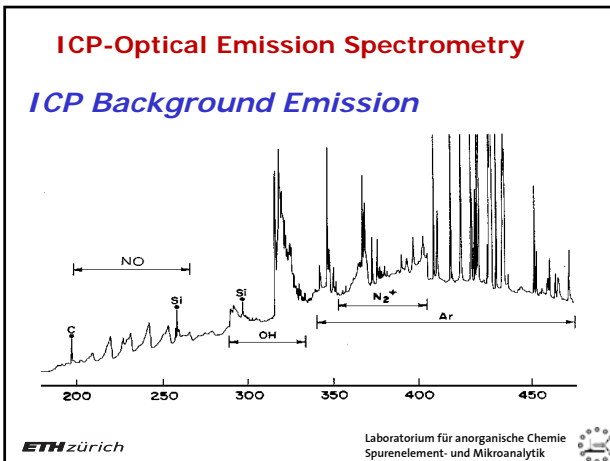


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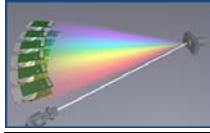




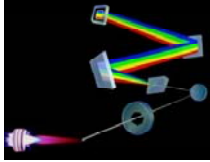
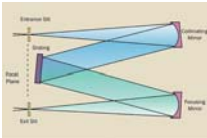
ICP-Optical Emission Spectrometry

High-resolution Optical Spectrometers required

$\Delta\lambda$: ≈ 10 s of pm
 Polychromators providing simultaneous access to the UV-VIS spectral range ($\approx 130/180 - 800$ nm)
 → Paschen Runge Setup
 → Echelle Configuration



Monochromators
 → Czerny Turner Configuration
 (barely used in current instruments)

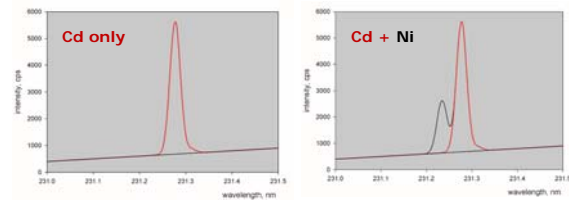


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ICP-Optical Emission Spectrometry

High-resolution Optical Spectrometers required



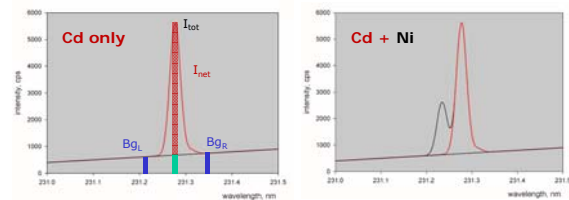
Identification and Correction of Spectral Interferences

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ICP-Optical Emission Spectrometry

Simultaneous Spectrometers advantageous



- Correlated acquisition allows for better compensation of fluctuating baseline and interference signals.
- Internal Standard(s) can be monitored simultaneously

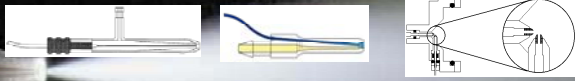
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ICP-Optical Emission Spectrometry

Sample Introduction Methods

Pneumatic Nebulizer (aerosol generation)



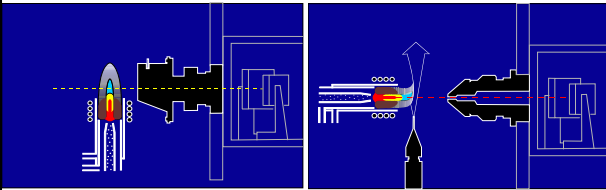
Spray Chamber
(removal of large droplets)

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ICP-Optical Emission Spectrometry

Plasma View: Radial vs. Axial



Robust against Matrix
Simpler Setup
Lower Analyte Sensitivity

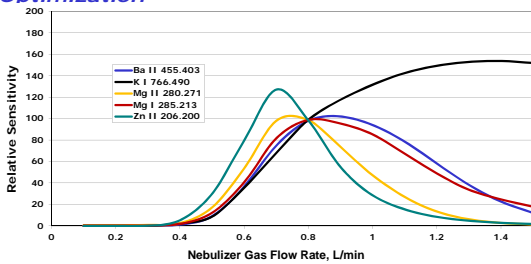
≈ 10x higher Sensitivity
Lower Background
Better LODs
Stronger Matrix Dependence

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ICP-Optical Emission Spectrometry

Optimization



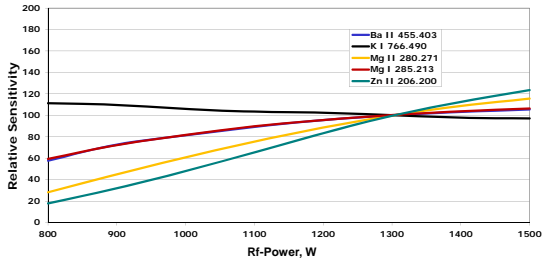
I: Atom Line, II: Ion Line

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ICP-Optical Emission Spectrometry

Optimization



I: Atom Line, II: Ion Line

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ICP-Optical Emission Spectrometry

Effects of Increasing Plasma Temperature:

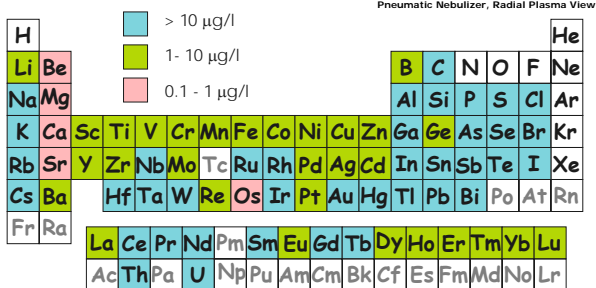
- More efficient vaporization and excitation
 - Higher Analyte Signal
 - Less Matrix Effects
 - More Potential Spectral Interferences
- Increasing degree of ionization
 - More intense Ion Lines
 - Less intense Atom Lines
 - More Potential Spectral Interferences
- Increasing background emission
 - Lower Signal/Noise
- Mechanical Stress / Erosion of the Torch

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ICP-Optical Emission Spectrometry

Typical Limits of Detection



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ICP-Optical Emission Spectrometry

Limitations:

- Sample Composition can Influence Emission Signal
 - More pronounced with Axial View
 - Enhancement and Suppression can occur
- Line-rich Spectra from many Transition Elements
 - Interference Control Mandatory
 - Background Signals may vary between Samples
- Deposition of dissolved Solids can cause Memory Effects
 - Long Rinse Times
 - Application-Specific Sample Introduction Systems

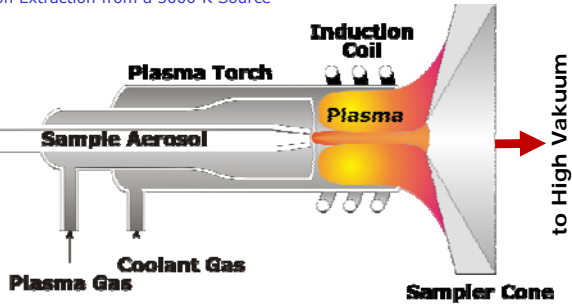
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ICP-Mass Spectrometry

Ion Extraction from a 5000 K Source



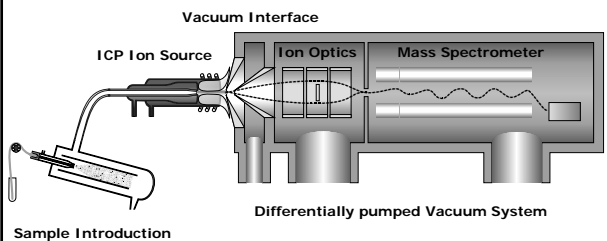
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ICP-Mass Spectrometry

Setup



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ICP-Mass Spectrometry

Why ICPMS?

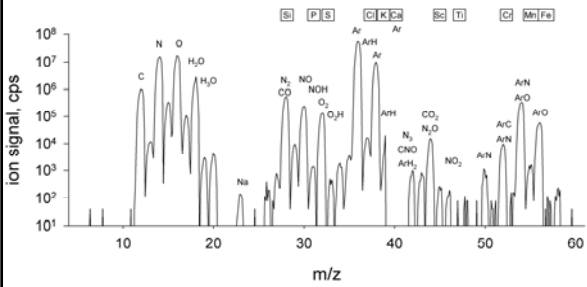
- Eventually lowest instrumental background
- High ion yield for most elements
 - Low Limits of Detection
- Less, and easier to predict spectral interferences
- Isotope Information
- Various Sample Introduction Methods
- Up to 12 Orders of Magnitude Linear Dynamic Range

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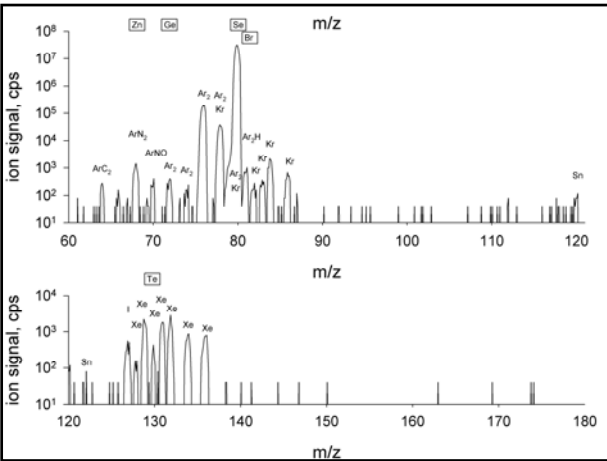
ICP-Mass Spectrometry

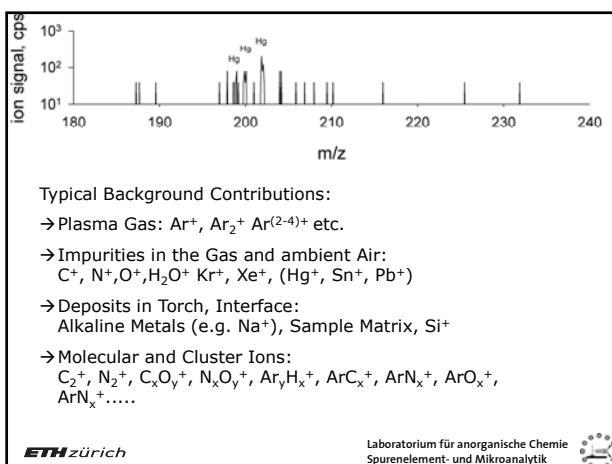
Intrumental Background

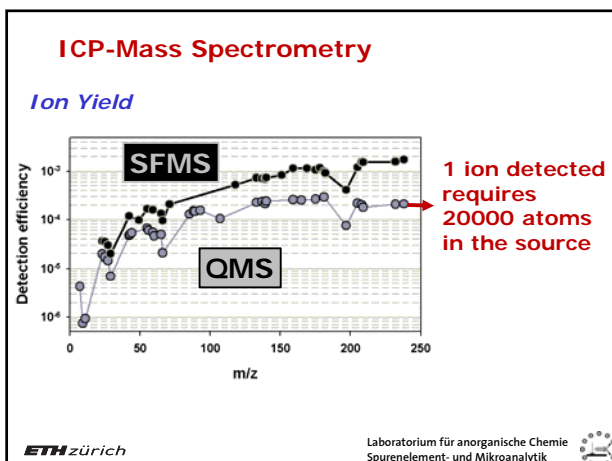


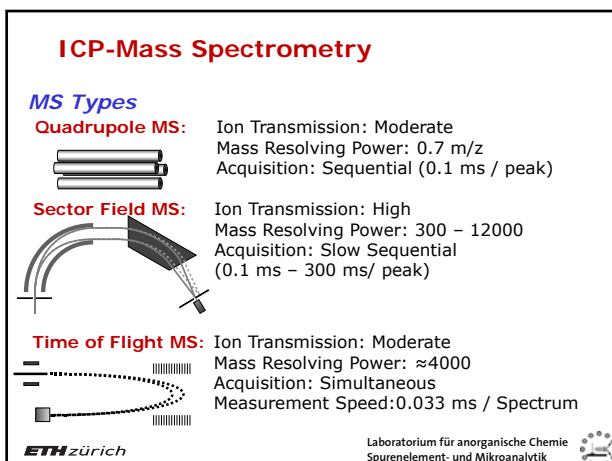
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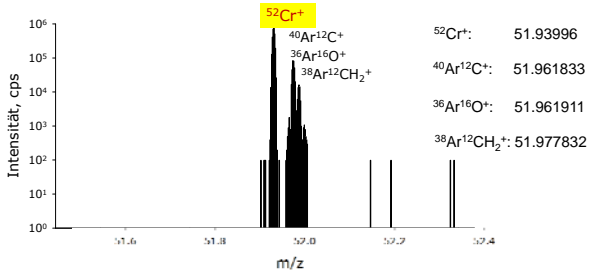




ICP-Mass Spectrometry

High-Resolution Advantage

At least 5 different Species in a 1 m/z Mass Window



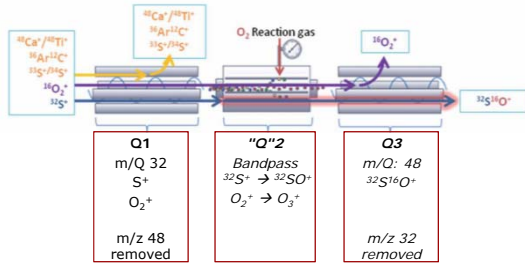
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ICP-Mass Spectrometry

"High Mass Resolution" with Quadrupole MS

Chemical manipulation of the Ion Beam Composition:
Ion-Molecule Reactions, MS-MS mode



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ICP-Mass Spectrometry

Limitations

Matrix Effects are more severe:

- > Spectral Interferences
 - o Limits the Choice of Solvents
(H₂SO₄, H₃PO₄ should be avoided, HCl only when absolutely needed)
- > Deposits at Vacuum Interface can cause Instrument Drift
 - o Limits the Content of dissolved Solids
(1 g/L generally considered maximum)
- > «Space Charge» Effects from high Ion Currents
- > Changes in Ion Yield:
 - o Signal Suppression (e.g. Easily Ionized Elements)
 - o Signal Enhancement (e.g. changes in Space Charge)

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ICP-Mass Spectrometry

Typical Limits of Detection

Pneumatic Nebulizer, Quadrupole MS, Aqueous Samples

H																	He
Li	Be									B	C	N	O	F	Ne		
Na	Mg									Al	Si	P	S	Cl	Ar		
K	Ca	Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn	Ga	Ge	As	Se	Br	Kr
Rb	Sr	Y	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Ag	Cd	In	Sn	Sb	Te	I	Xe
Cs	Ba		Hf	Ta	W	Re	Os	Ir	Pt	Au	Hg	Tl	Pb	Bi	Po	At	Rn
Fr	Ra																
		La	Ce	Pr	Nd	Pm	Sm	Eu	Gd	Tb	Dy	Ho	Er	Tm	Yb	Lu	
		Ac	Th	Pa	U	Np	Pu	Am	Cm	Bk	Cf	Es	Fm	Md	No	Lr	

■ > 100 ng/L
■ > 1 ng/L
■ < 1 ng/L

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ICP-based Spectrometries

Characteristics

	OES	MS
Sample Volume	mL	µL-mL
Analysis Time	seconds	seconds
Matrix Tolerance ¹	high (%)	low (<0.1%)
Calibration ²	Ext./Std.Add.	Ext./Std.Add./ID
Repeatability ³	1-5%	1-5%
Dynamic Range	10 ⁶	10 ⁹ -10 ¹²

1: Values in Brackets indicate the tolerable Dissolved Solids Content
 2: Ext: External Calibration, Std.Add.: Standard Addition, ID: Isotope Dilution
 3: For Concentrations > 50×LOD

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Calibration

Methods

- > External Calibration
 - o Synthetic Solutions of the Analyte(s)
 - o Matrix-Matching may be required
 - o Internal Standards recommended (ICP-based methods)
- > Standard Addition
 - o Addition of Synthetic Solutions of the Analyte(s) to Aliquots of the Sample
 - o Ideal Matrix-Matching
 - o Time-consuming
- > Isotope Dilution
 - o Only with MS

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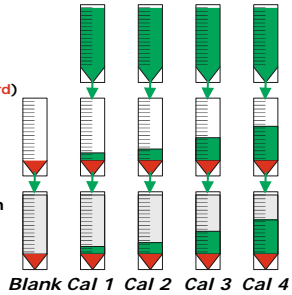
Calibration

External Calibration

Known Amounts of Primary Standard(s) are filled into clean vials (eventually with an Internal Standard)
Ideal to use different Supplies

Dilution to desired Concentration using a suitable Solvent

Mix!



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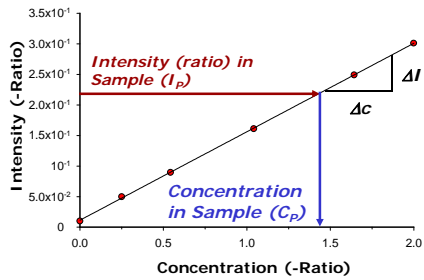
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Calibration

External Calibration

$$C_p = \frac{I_p - I_0}{S}$$

$$S = \frac{\Delta I}{\Delta C}$$



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Calibration

External Calibration

- + Simple
- + Practically all Elements accessible
- + Practically all Sample Types accessible
- + Internal Standard can be easily included
- + Concentration range only limited by Solubilities

- Spectral Interferences need to be absent or corrected for
- Matrix Effects and Instrument Drift have to be monitored and corrected for

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Calibration

Standard Additions

Sample splitted to identical Aliquots
→ minimum 3, ideal 5

Addition of known amounts of a Standard Solution

Dilution to desired Concentration using a suitable Solvent

Mix!

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Calibration

Standard Additions

$C_p = \frac{I_0}{S}$

$S = \frac{\Delta I}{\Delta C}$

Round Robin Test
IMEP-16, IRMM
Pb in Wine

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Calibration

Standard Additions

IMEP-16: Pb in wine
Certified value : $27.18 \pm 0.33 \mu\text{g}\cdot\text{L}^{-1}$ [$U=k\cdot u_c$ ($k=2$)]

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Calibration

Standard Additions

+ Matrix Effects can be eliminated

- Spectral Interferences need to be absent or corrected for
- Labour intensive
- Approximate Concentration needs to be known
- Long measurement times
- Instrument Drift is not compensated for (may require another internal Standard)

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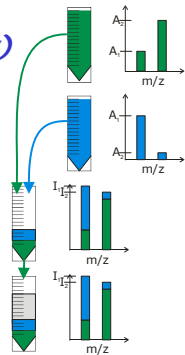
Isotope Dilution (MS only)

Unknown Sample with «Natural»
Isotope Ratio
Isotopically enriched Standard

Aliquots of exactly known
amount are mixed

Dilution to desired Concentration
using a suitable Solvent

Mix!



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Calibration

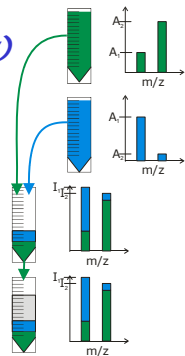
Isotope Dilution (MS only)

$$R_P = \frac{A_{1P}}{A_{2P}}$$

$$R_S = \frac{A_{1S}}{A_{2S}}$$

$$R_M = \frac{I_{1M}}{I_{2M}}$$

$$C_P = C_S \times \frac{V_S}{V_P} \times \frac{R_S - R_M}{R_M - R_P} \times \frac{A_{1P}}{A_{1S}}$$



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Calibration

Isotope Dilution (MS only)

- + "Goldstandard" of Calibration
- + Optimum internal Standard
- + Ideal Compensation of Matrix Effects
- + SI-Traceable
- High experimental Effort
- Not Applicable with Mono-Isotopic Elements
- Spectral Interferences must be absent or corrected for
- Enriched Standards not readily available
- Instrument Drift not easily compensated for

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Sample Preparation

Aqueous Solutions

- Stabilization of the Elements in Solution
 - Usually Acidification with HNO_3
(Saturation of Cation-Exchange positions of the Container and Sample Introduction)
 - Specific Stabilizers may be required, depending on Chemistry
- Dilution to suitable matrix concentration
 - Depends on Analytical Method
- Addition of Internal Standard(s)

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Sample Preparation

Organic Solvents

- High variability of Viscosity, Surface Tension and Evaporation Rates causes issues with Nebulization
 - Sample uptake Rate needs to be controlled
- Calibration Standards not readily available
- ICP methods are less tolerant to Organic Solvents
 - Digestion
 - Addition of trace amounts O_2 to the ICP to avoid Soot formation

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Sample Preparation

Solids

Conventional Methods require Digestion using a suitable Reagents

Most frequently an Acid Cocktail

→ HNO_3 (HCl, HF if necessary; H_2SO_4 should be avoided)

Suitable Digestion Methods are available for most Materials

→ Reagents and Digestion Vials need to be of sufficient Purity

Direct Solids analysis using Laser Ablation ICPMS or Solid Sampling GFAAS

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Sample Preparation

Common Digestion Methods

Microwave assisted High Pressure, High Temperature Digestion

→ Temperature up to 200 °C, Pressure up to 200 bar



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Sample Preparation

Common Digestion Methods

Oven-heated High Temperature, High Pressure Systems



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Sample Preparation

Common Digestion Methods

All Material in Contact with Sample and Reagents must be Inert and Clean

- Mostly Fluoropolymers: PTFE, PFA, etc.
- (Quartz)

Evolution of Gas (e.g. Oxidation of Organic Material) can lead to sudden Pressure Jump

- Sample loss via the Safety Relief Valve / Rupture Disk
- Potential Safety Hazard

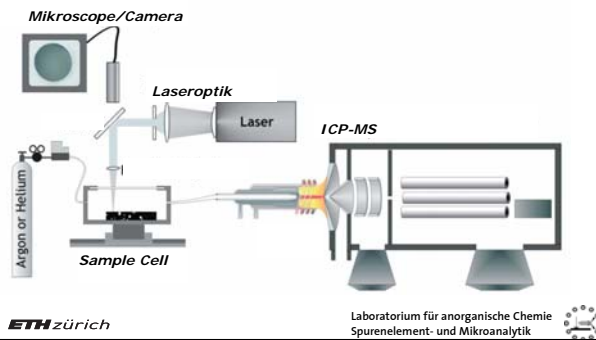
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Sample Preparation

Direct Solid Sampling with LA



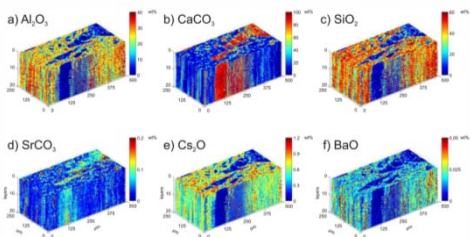
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Sample Preparation

Direct Solid Sampling with LA



2-D and 3-D Element Distributions at 5 µm Spatial Resolution

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Method Development

Considerations

Accuracy of the Analysis

Concentration Range

Techniques used

Reproducibility

Sample Throughput

Cost

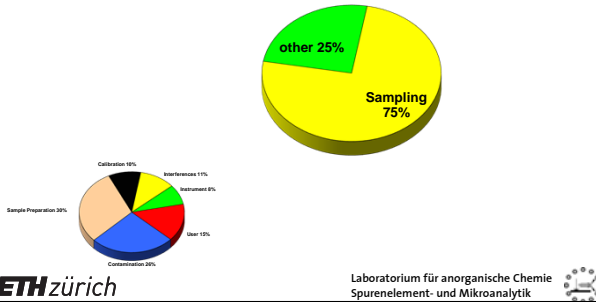
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Method Development

Sources of Error in Trace Element Analysis



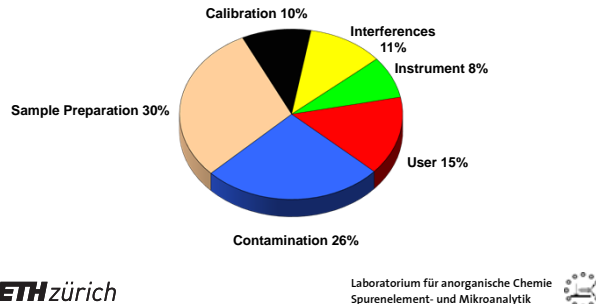
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Method Development

Sources of Error in Trace Element Analysis



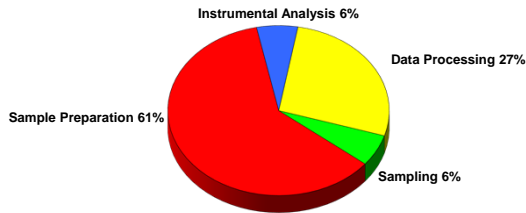
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Method Development

Time Spent



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Method Development

Validation

Ensuring Representative Analytical Results

- **Validate Representativeness of Subsamples**
Sampling Theory
- **Validate Accuracy of Calibration**
Recovery Check Analyses
Serial Dilution
Matrix Evaluation
Analysis of Certified Reference Materials
- **Evaluate Reproducibility of Results**
Duplicate Analyses (randomized)
Repeated Calibration

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